



Product Manual

AMPIGENE[®] Taq DNA Polymerase

Catalog #: ENZ-PRT100

ENZ-PRT100-0500 for 500 units



Product Manual

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Please read entire booklet before proceeding with the assay.



Carefully note the handling and storage conditions.



Please contact Enzo Life Sciences Technical Support if necessary.

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DESCRIPTION

AMPIGENE[®] Taq DNA Polymerase uses the latest developments in polymerase technology and buffer chemistry to enhance PCR speed, yield and specificity. The enzyme and buffer system allow for superior PCR performance on complex templates such as mammalian genomic DNA.

AMPIGENE[®] Taq is a robust enzyme for all your everyday PCR applications including genotyping, screening and library construction. AMPIGENE[®] Taq DNA Polymerase can perform consistently well on a broad range of templates (including both GC and AT rich).

AMPIGENE[®] Taq DNA Polymerase has an error rate of approximately 1 error per 2.0×10^5 nucleotides incorporated. PCR products generated with AMPIGENE[®] Taq DNA Polymerase are A-tailed and may be cloned into TA cloning vectors.

Component	500 units
AMPIGENE [®] Taq DNA polymerase (5 u/μL)	1 x 100 μL
5x AMPIGENE [®] reaction buffer	4 x 1 mL



Protect from prolonged exposure to light.

SHIPPING AND STORAGE

On arrival the kit should be stored at -20°C. Avoid prolonged exposure to light. If stored correctly the kit will retain full activity for 12 months. The kit can be stored at 4°C for 1 month. The kit can go through 30 freeze/thaw cycles with no loss of activity.



IMPORTANT CONSIDERATIONS

5x AMPIGENE[®] Reaction buffer: The 5x reaction buffer contains 15 mM MgCl₂, 5 mM dNTPs, enhancers and stabilizers. It is not recommended to add further PCR enhancers or MgCl₂ to the reaction. The buffer composition has been optimized to maximize PCR success rates.

Template: For eukaryotic DNA use between 5 ng and 500 ng per reaction, for cDNA use below 100 ng per reaction.

Primers: Primers should have a predicted melting temperature of around 60°C. The final primer concentration in the reaction should be between 0.2 μM and 0.6 μM.

Annealing: We recommend performing a temperature gradient to experimentally determine the optimal annealing temperature. Alternatively, we recommend a 55°C annealing temperature then increase in 2°C increments if non-specific products are present.

Extension: Optimal extension is achieved at 72°C. The optimal extension time is dependent on amplicon length and complexity of template. 15 seconds per kilobase (kb) is recommended for amplification from eukaryotic DNA for amplicons between 1kb and 6kb. For shorter amplicons a 1 second extension is sufficient.

REACTION SETUP

1. Prepare a master mix based on the following table:

Reagent	50 μ L reaction	Final concentration	Notes
5x AMPIGENE [®] Reaction buffer	10.0 μ L	1x	
Forward primer (10 μ M)	2.0 μ L	400 nM	See above for optimal primer design
Reverse primer (10 μ M)	2.0 μ L	400 nM	
Template DNA	<100 ng cDNA, <500 ng genomic	variable	See above for template considerations
AMPIGENE [®] Taq DNA polymerase (5 u/ μ L)	0.25 μ L - 1.0 μ L		
PCR grade dH ₂ O	Up to 50 μ L final volume		

2. Cycle using conditions based on the following table:

Cycles	Temperature	Time	Notes
1	95°C	1 min	Initial denaturation
40	95°C	15 seconds	Denaturation
	55°C to 65°C	15 seconds	Anneal
	72°C	1-90 seconds	Extension (15 seconds per kb)



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NOTES



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