

Cyclic GMP Complete

Catalog #: ADI-901-164

5 x 96-well Kit

For use with cells, tissue, saliva, serum, plasma, urine and culture supernates

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Product Manual

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All reagents, except standard and conjugate, should be stored at 4°C. Store standard and conjugate at -20°C



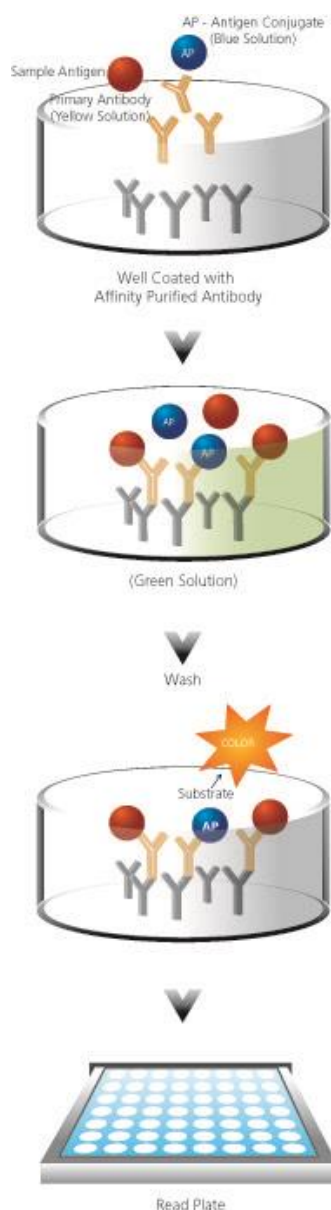
For proper performance, use the insert provided with each individual kit received.



Check our website for additional protocols, technical notes and FAQs.

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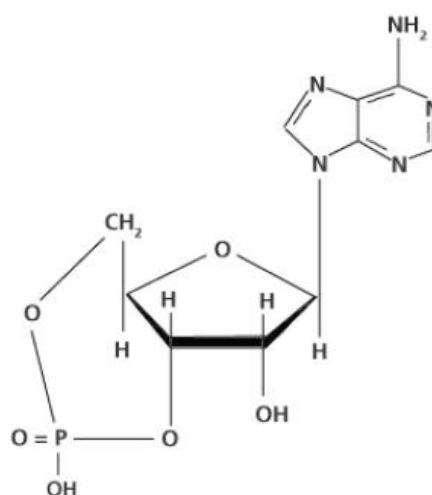
INTRODUCTION



The cGMP complete Enzyme-linked Immunosorbent Assay (ELISA) kit is a competitive immunoassay for the quantitative determination of cyclic GMP in cells and tissue treated with 0.1M HCl, in addition to culture supernatants, saliva, and serum. The optional acetylated assay format provides an approximate 10-fold increase in sensitivity and is ideal for samples with extremely low levels of cGMP. If expected levels of cGMP are unknown, the investigator may evaluate a few samples in the non-acetylated format in order to determine if higher sensitivity is required.

Guanosine 3',5'-cyclic monophosphate (cyclic GMP; cGMP) was identified in 1963¹. It has been shown to be present at levels typically 10-100 fold lower than cAMP in most tissues and is formed by the action of the enzyme guanylate cyclase on GTP. It is involved in a number of important biological reactions. Some hormones, such as acetylcholine, insulin, and oxytocin, as well as certain other chemicals like serotonin and histamine cause an increase in cGMP levels^{2,3}. Stimulators of guanylate cyclase such as the vasodilators nitroprusside, nitroglycerin, sodium nitrate, and nitric oxide (NO) also stimulate cGMP levels⁴. Peptides, such as atrial natriuretic peptide (ANP) that relax smooth muscle also increase cGMP concentrations⁵. cGMP has been confirmed as a second messenger for ANP⁶. NO can be synthesized from L-arginine and diffuse through cell membranes^{7,8}. The interaction of NO with guanylate cyclase allows cGMP to act as a third messenger in some cells⁹.

cyclic GMP



PRINCIPLE

1. Standards and samples are added to wells coated with a GxR IgG antibody. A blue solution of cGMP conjugated to alkaline phosphatase is then added, followed by a yellow solution of rabbit polyclonal antibody to cGMP.
2. During a simultaneous incubation at room temperature the antibody binds, in a competitive manner, the cGMP in the sample or conjugate. The plate is washed, leaving only bound cGMP.
3. pNpp substrate solution is added. The substrate generates a yellow color when catalyzed by the alkaline phosphatase on the cGMP conjugate.
4. Stop solution is added. The yellow color is read at 405nm. The amount of signal is indirectly proportional to the amount of cGMP in the sample.



Do not mix components from different kit lots or use reagents beyond the expiration date of the kit.



HCl is caustic. Keep tightly capped.



The standard should be handled with care due to the known and unknown effects of the antigen.



Triethylamine and acetic anhydride are lachrymators. Caution—corrosive, flammable, and harmful vapor.



Avoid contamination by endogenous alkaline phosphatase. Do not expose reagents or supplies to bare skin.



Activity of conjugate is affected by concentrations of chelators > 10mM (such as EDTA and EGTA).



Stop solution is caustic. Keep tightly capped.

MATERIALS SUPPLIED

1. **Assay Buffer 2 Concentrate**
100 mL, Product No. 80-0074
Sodium acetate buffer containing proteins and sodium azide
2. **0.1M HCl**
110 mL, Product No. 80-0082
0.1M hydrochloric acid in water
3. **cyclic GMP Standard**
3 x 0.5 mL, Product No. 80-0153
A solution of 5,000 pmol/mL cGMP
4. **Acetylation Kit**
2 x 2 vials, Product No. 950-001
 - a. **Triethylamine**
2 x 2 mL, Product No. 80-0063
 - b. **Acetic Anhydride**
2 x 1 mL, Product No. 80-0064
5. **Goat anti-Rabbit IgG Microtiter Plate**
Five plates of 96 wells, Product No. 80-0060
A clear plate of break-apart strips coated with a goat anti-rabbit polyclonal antibody
6. **Neutralizing Reagent**
27 mL, Product No. 80-1476
7. **cGMP Antibody**
25 mL, Product No. 80-0161
A yellow solution of rabbit polyclonal antibody to cGMP
8. **cGMP Conjugate**
25 mL, Product No. 80-0159
A blue solution of cGMP conjugated to alkaline phosphatase
9. **Wash Buffer Concentrate**
100 mL, Product No. 80-1287
Tris buffered saline containing detergents
10. **pNpp Substrate**
100 mL, Product No. 80-0076
A solution of p-nitrophenyl phosphate
11. **Stop Solution**
30 mL, Product No. 80-0248
A solution of trisodium phosphate in water
12. **cGMP Complete Assay Layout Sheet**
1 each, Product No. 30-0247
13. **Plate Sealer**
5 each, Product No. 30-0012



Reagents require separate storage conditions.

STORAGE

All components of this kit, **except the Conjugate and Standard**, are stable at 4°C until the kit's expiration date. Upon receipt, store the Conjugate and Standard at -20°C. Recommended storage temperatures do not necessarily reflect shipping conditions.

MATERIALS NEEDED BUT NOT SUPPLIED

1. Deionized or distilled water
2. Precision pipets for volumes between 5 µL and 1,000 µL
3. Repeater pipet for dispensing 50 µL and 200 µL
4. Disposable beakers for diluting buffer concentrates
5. Graduated cylinders
6. Microplate shaker
7. Lint-free paper toweling for blotting
8. Microplate reader capable of reading at 405 nm



Glass or polypropylene tubes may be used for

REAGENT PREPARATION

1. Assay Buffer 2

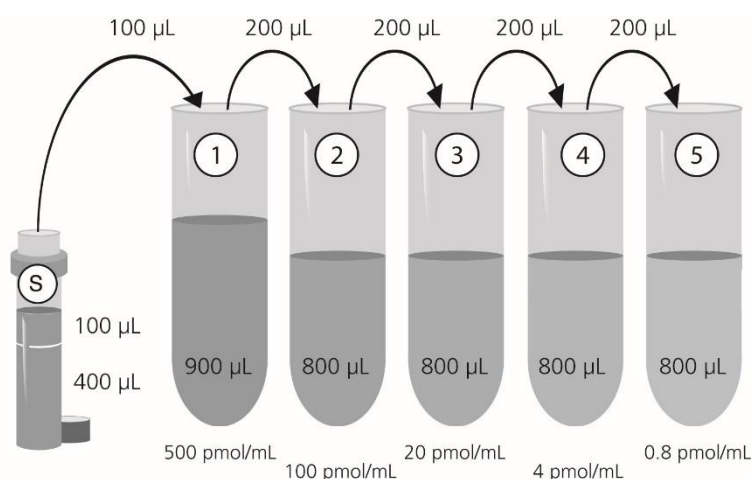
Prepare Assay Buffer 2 by diluting 50mL of the supplied 2x concentrate with 50mL of deionized water. Discard unused buffer or add up to 0.09% sodium azide (w/v) for storage.

2. Wash Buffer

Prepare the wash buffer by diluting 5 mL of the supplied Wash Buffer Concentrate with 95 mL of deionized water. This can be stored at room temperature until the kit expiration, or for 3 months, whichever is earlier.

3. cGMP Standard, non-acetylated format

Three diluent options are available for the preparation of the standard curve, NSB, and Bo wells. For cell lysates and tissue samples prepared in 0.1M HCl, use the supplied 0.1M HCl as the standard diluent. Use Assay Buffer 2 for the standard diluent when the sample is serum, plasma, urine, or saliva. For culture supernatants, use the same non-conditioned media for the standard diluent.



Allow the 5,000 pmol/mL standard stock to warm to room temperature. Label five 12mm x 75mm tubes #1 through #5. Pipet 900 µL of the appropriate sample diluent into tube #1. Pipet 800 µL of the appropriate sample diluent into tubes #2 through #5. Add 100 µL of the 5,000 pmol/mL standard stock into tube #1 and vortex thoroughly. Add 200 µL of tube #1 to tube #2 and vortex thoroughly. Add 200 µL of tube #2 to tube #3 and vortex thoroughly. Continue this for tubes #4 through #5.

Diluted standards should be used within 60 minutes of preparation. The concentrations of cGMP in the tubes are labeled above.

4. Acetylation Reagent (optional)



Triethylamine and acetic anhydride are lachrymators. Caution-

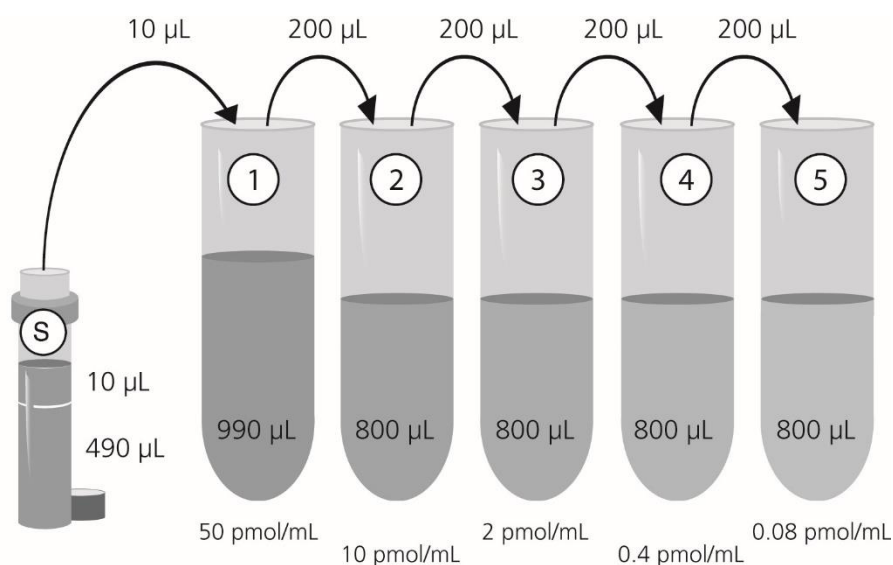


Glass or polypropylene tubes may be used for standard preparation. Avoid polystyrene.

Prepare the Acetylating Reagent by adding 0.5 mL of Acetic Anhydride to 1 mL of Triethylamine. Note that this volume is sufficient to add to 30 mL of diluted standards and samples. Use the prepared reagent within 60 minutes of preparation. Discard any unused portion of the Acetylating Reagent.

5. cGMP Standard, acetylated format (optional)

Three diluent options are available for the preparation of the standard curve, NSB, and Bo wells. For cell lysates and tissue samples prepared in 0.1M HCl, use the supplied 0.1M HCl as the standard diluent. Use Assay Buffer 2 for the standard diluent when the sample is serum or saliva. For culture supernatants, use the same non-conditioned media for the standard diluent.



Allow the 5,000 pmol/mL standard stock to warm to room temperature. Label five 12mm x 75mm tubes #1 through #5. Pipet 990 µL of the appropriate sample diluent into tube #1. Pipet 800 µL of the appropriate sample diluent into tubes #2 through #5. Add 10 µL of the 5,000 pmol/mL standard stock into tube #1 and vortex thoroughly. Add 200 µL of tube #1 to tube #2 and vortex thoroughly. Add 200 µL of tube #2 to tube #3 and vortex thoroughly. Continue this for tubes #4 through #5.

Acetylate all **standards and samples** by adding 10 µL of the Acetylating Reagent for each 200 µL of the standard or sample. Add the Acetylating Reagent directly to the diluted standard or sample and vortex immediately after the addition of the Acetylating Reagent.

Label one 12mm x 75mm tube as the Bo/NSB tube. Pipet 1 mL of the appropriate standard diluent into this tube. Add 50 μ L of the Acetylating Reagent to the Bo/NSB tube and use in Steps 2 and 3 of the Assay Procedure.

The acetylated standards should be used within 30 minutes of preparation. The concentrations of cGMP in the tubes are labeled above.

SAMPLE HANDLING



If buffers other than those provided are used in the assay, the end-user must determine the appropriate dilution and assay validation.



Samples must be stored at or below -20°C to avoid loss of bioactive analyte. Repeated freeze/thaw cycles should be avoided.

Treatment of cells and tissue with the supplied 0.1M HCl will stop endogenous phosphodiesterase activity and allow for the direct measurement of these samples in the assay without evaporation or further processing. Recommended treatment protocols follow. Samples containing rabbit IgG will interfere with the assay. EDTA plasma may precipitate during acetylation.

Biological fluids, such as serum, plasma, urine, and saliva, should be diluted in Assay Buffer 2 and run directly in the assay. Culture Supernatants should also be diluted in Assay Buffer 2 and the same non-conditioned media diluted with the assay buffer should be used as the standard diluent.

Please note that some samples may contain high levels of cGMP and additional dilution may be required. Samples with low levels of cGMP may be assayed in the acetylated format or the samples may be concentrated.

Protocol for Cell Lysates

The concentration of cells used must be optimized for the specific cell line and treatment conditions. Cells may be grown in typical containers such as Petri dishes, culture plates (e.g., 48-well, 12-well, or 96-well), culture flasks, etc. Some cells are particularly hardy (e.g., bacteria) and may require the addition of 0.1 to 1% Triton x-100 to the 0.1M HCl for enhanced lysis. If Triton x-100 is added to samples it should also be added to the standard dilution as a modest increase in optical density may occur.

1. Pellet **suspension cells** and aspirate the media. Treat cells with 0.1M HCl. A general starting concentration of 1×10^6 cells per mL of 0.1M HCl is recommended. Remove the media from **adherent cells** and add enough 0.1M HCl to cover the bottom of the plate. Avoid over-diluting the sample with an excessive volume of HCl. Please note that the culture media may be saved and assayed separately, if desired.
2. Incubate the cells in 0.1M HCl for 10 minutes at room temperature. Inspect the cells under a microscope to ensure uniform lysis. Continue incubating for an additional 10 minutes, if necessary.

3. Centrifuge $\geq 600 \times g$ to pellet the cellular debris.
4. The supernatant may be assayed immediately or stored frozen for later analysis.

Note: Standards must be diluted in 0.1 M HCl and Neutralizing Reagent used.

Protocol for Tissue Samples

1. After collection, tissue samples should be flash frozen in liquid nitrogen. If analysis cannot be carried out immediately, store tissue at -80°C .
2. Grind frozen tissue to a fine powder under liquid nitrogen in a stainless steel mortar.
3. When liquid nitrogen has evaporated, weigh the frozen tissue and homogenize in 10 volumes of 0.1M HCl (e.g., 0.1 g of tissue should be homogenized in 1 mL of 0.1M HCl).
4. Centrifuge $\geq 600 \times g$ to pellet the debris (~10 minutes).
5. The supernatant may be further diluted in the 0.1M HCl provided and run directly in the assay or stored frozen for later analysis.

Note: standards must be diluted in 0.1 M HCl and Neutralizing Reagent used.


SAMPLE RECOVERIES


cGMP standard was spiked into the following matrices diluted with Assay Buffer 2 and measured in the kit. The results were as follows:


Sample	% Recovery	Recommended Dilution, Acetylated Format	Recommended Dilution, Non-Acetylated Format
Tissue Culture Media	101.7%	Undiluted	1:100
Human Saliva	102.9%		1:10
Human Serum	101.3%	Not recommended	≥1:10
Human Heparin Plasma	104.4%		1:10
Human EDTA Plasma	115.0%	≥1:2	≥1:10
Human Urine	97.7%		≥1:100


0.1 M HCl should not be used to dilute culture supernates, serum, plasma, urine or saliva samples.


ASSAY PROCEDURE

 Bring all reagents to room temperature for at least 30 minutes prior to opening.

 All standards and samples should be run in duplicate.

 Pre-rinse each pipet tip with reagent. Use fresh pipet tips for each sample, standard, and reagent.

 Pipet the reagents to the sides of the wells to avoid possible contamination.

 Prior to the addition of substrate, ensure there is no residual wash buffer in the wells. Remaining wash buffer may cause variation in assay results.

Refer to the Assay Layout Sheet to determine the number of wells to be used. Remove the wells not needed for the assay and return them, with the desiccant, to the mylar bag and seal. Store unused wells at 4°C.

Note: If the **acetylated format** of the assay is to be run, all standards, samples, and the diluent for the NSB and Bo wells must be acetylated as per the instructions in the Reagent Preparation section. Acetylated standards and samples must be used within 30 minutes.

1. If using samples prepared in 0.1M HCl, pipet 50 μ L of Neutralizing Reagent into each well except the Total Activity (TA) and Blank wells. Do not add Neutralizing Reagent for the other sample diluent options.
2. Pipet 100 μ L of the appropriate standard diluent (Assay Buffer 2, 0.1M HCl, or non-conditioned culture media) into the NSB (non-specific binding) and Bo (0 pmol/mL standard) wells.
3. Add 50 μ L of the appropriate standard diluent to the NSB wells.
4. Pipet 100 μ L of Standards #1 through #5 to the bottom of the appropriate wells.
5. Pipet 100 μ L of the samples to the bottom of the appropriate wells.
6. Pipet 50 μ L of the blue conjugate into each well except the TA and Blank wells.
7. Pipet 50 μ L of the yellow antibody into each well except the Blank, TA, and NSB wells.
Note: Every well used should be green in color except the NSB wells which should be blue. The Blank and TA wells are empty at this point and have no color.
8. Seal the plate. Incubate for 2 hours on a plate shaker (~500 rpm) at room temperature.
9. Empty the contents of the wells and wash by adding 400 μ L of wash buffer to every well. Repeat 2 more times for a total of 3 washes. After the final wash, empty or aspirate the wells by inverting the plate and firmly tapping it on a lint-free paper towel to remove any remaining wash buffer.
10. Pipet 5 μ L of the blue conjugate to the TA wells.
11. Add 200 μ L of the substrate solution into each well.
12. Incubate for 1 hour at room temperature without shaking.
13. Pipet 50 μ L stop solution into each well.

14. After blanking the plate reader against the substrate blank, read optical density at 405 nm. If plate reader is not capable of adjusting for the blank, manually subtract the mean OD of the substrate blank from all readings.

CALCULATION OF RESULTS

Several options are available for the calculation of the concentration of cGMP in samples. We recommend that the data be handled by an immunoassay software package such as Assay Blaster Data Analysis Software (Product no. ADI-28-0002) utilizing a 4 parameter logistic curve fitting program. If data reduction software is not readily available, the concentrations can be calculated as follows:

1. Calculate the average net OD for each standard and sample by subtracting the average NSB OD from the average OD for each standard and sample.

$$\text{Average Net OD} = \text{Average OD} - \text{Average NSB OD}$$

2. Calculate the binding of each pair of standard wells as a percentage of the maximum binding wells (B_0), using the following formula:

$$\text{Percent Bound} = \frac{\text{Net OD}}{\text{Net } B_0 \text{ OD}} \times 100$$

3. Plot the Percent Bound (B/B_0) versus concentration of cGMP for the standards. Approximate a straight line through the points. The concentration of cGMP of the unknowns can be determined by interpolation.

Samples with concentrations outside of the standard curve range will need to be re-analyzed using a different dilution.

To normalize for protein content, divide the resulting picomole per mL determinations (pmol/mL) by the total protein concentration (mg/mL) in each sample. This is expressed as pmol cGMP per mg of total protein.



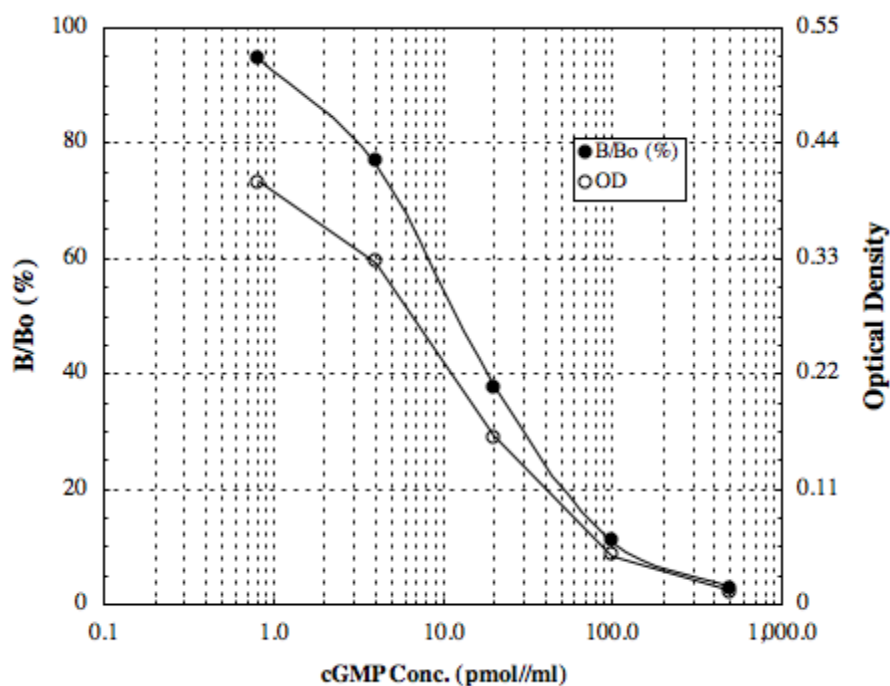
Make sure to multiply sample concentrations by the dilution factor used during sample preparation.

TYPICAL RESULTS

The results shown below are for illustration only and should not be used to calculate results from another assay.

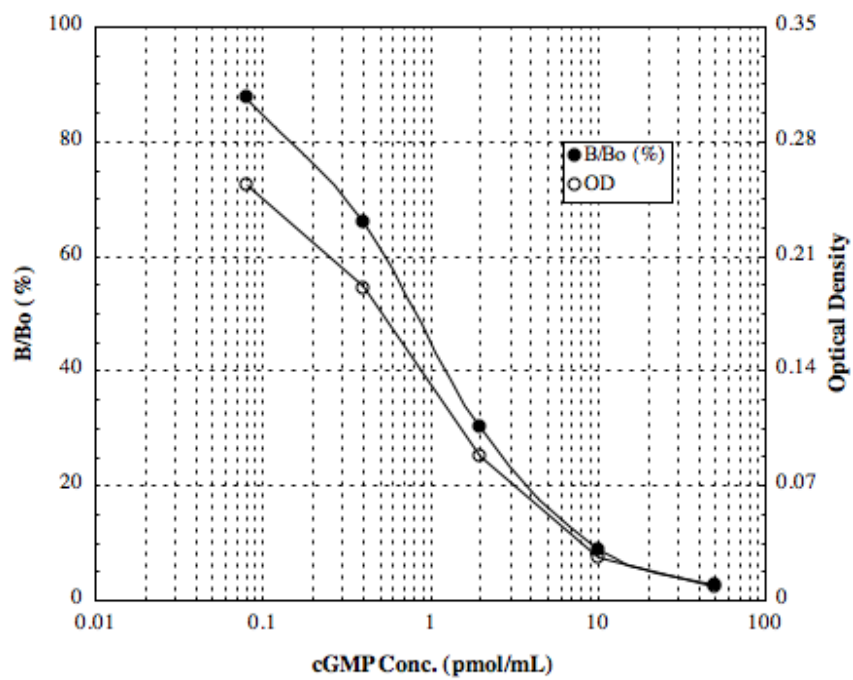
Non-acetylated assay format in Assay Buffer 2

Sample	Average Net OD	Percent Bound	cGMP (pmol/mL)
Blank (mean)	(0.086)	---	---
TA	0.251	---	---
NSB	-0.001	---	---
Bo	0.425	100%	0
S1	0.012	2.8%	500
S2	0.049	11.5%	100
S3	0.160	37.7%	20
S4	0.327	76.9%	4
S5	0.403	94.8%	0.8
Unknown 1	0.087	20.9%	47
Unknown 2	0.367	86.4%	2.2



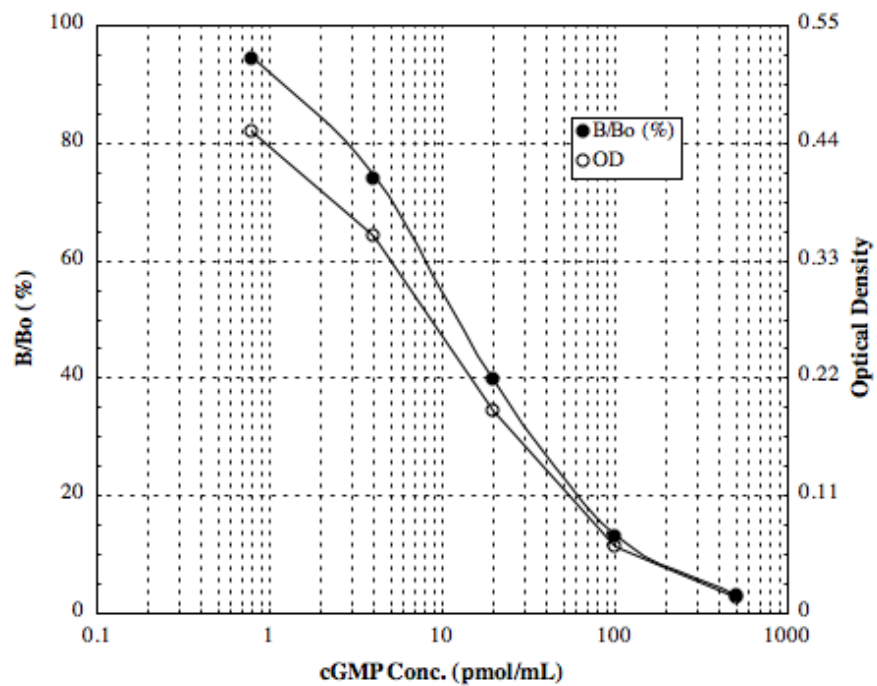
Acetylated assay format in Assay Buffer 2

Sample	Average Net OD	Percent Bound	cGMP (pmol/mL)
Blank (mean)	(0.091)	---	---
TA	0.254	---	---
NSB	-0.008	---	---
Bo	0.290	100%	0
S1	0.008	2.8%	50
S2	0.026	9.0%	10
S3	0.088	30.5%	2
S4	0.191	65.9%	0.5
S5	0.254	87.6%	0.08
Unknown 1	0.052	17.9%	4.2
Unknown 2	0.086	29.8%	2.1



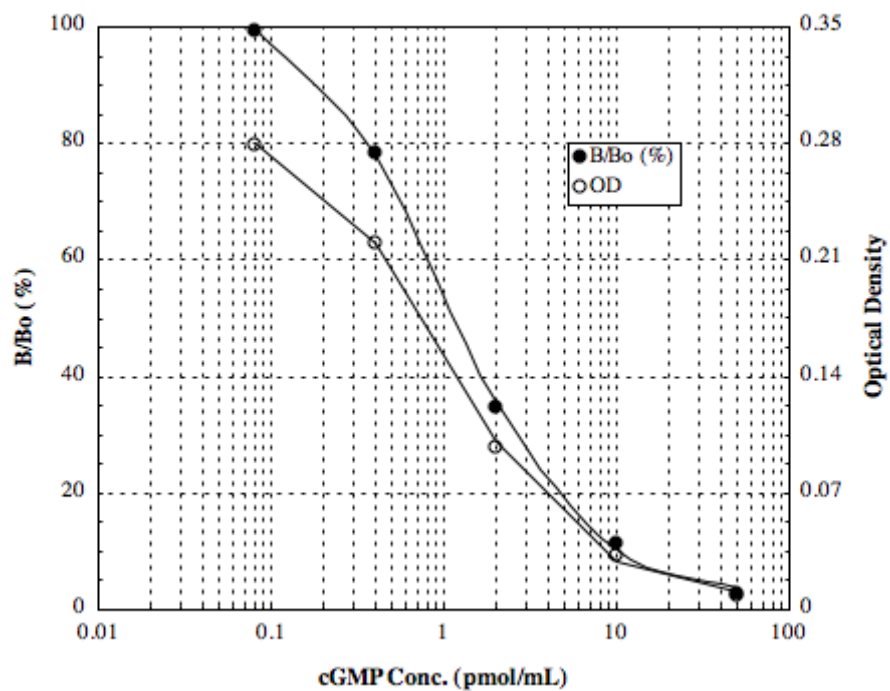
Non-acetylated assay format in 0.1 M HCl

Sample	Average Net OD	Percent Bound	cGMP (pmol/mL)
Blank (mean)	(0.094)	---	---
TA	0.298	---	---
NSB	0.000	---	---
Bo	0.478	100%	0
S1	0.015	3.1%	500
S2	0.062	13.0%	100
S3	0.190	39.7%	20
S4	0.354	74.1%	4
S5	0.451	94.4%	0.8
Unknown 1	0.145	30.5%	31
Unknown 2	0.344	71.9%	4.5



Acetylated assay format in 0.1 M HCl

Sample	Average Net OD	Percent Bound	cGMP (pmol/mL)
Blank (mean)	(0.123)	---	---
TA	0.335	---	---
NSB	-0.002	0%	---
Bo	0.281	100%	0
S1	0.008	2.8%	50
S2	0.032	11.4%	10
S3	0.098	34.9%	2
S4	0.220	78.3%	0.4
S5	0.279	99.3%	0.08
Unknown 1	0.023	8.2%	14
Unknown 2	0.055	19.9%	4.4



PERFORMANCE CHARACTERISTICS

Specificity

The cross reactivities for a number of related compounds were determined by diluting the cross reactants in the kit assay buffer at a concentration of ten times the high standard. These samples were then measured in the assay.

Compound	Cross Reactivity
cGMP	100%
GMP	<0.001%
GTP	<0.001%
cAMP	<0.001%
AMP	<0.001%
ATP	<0.001%
cUMP	<0.001%
CTP	<0.001%

Sensitivity

Assay Buffer 2

The sensitivity of the assay, defined as the concentration of cGMP measured at 2 standard deviations from the mean of 16 zeros along the standard curve, was determined to be 0.420 pmol/mL in the non-acetylated assay format and 0.043 pmol/mL in the acetylated assay format.

0.1M HCl

The sensitivity of the assay, defined as the concentration of cGMP measured at 2 standard deviations from the mean of 16 zeros along the standard curve, was determined to be 0.604 pmol/mL in the non-acetylated assay format and 0.059 pmol/mL in the acetylated assay format.

Linearity

A buffer sample containing cGMP was serially diluted 1:2 in the kit assay buffer and measured in the assay. The results are shown in the table below.

Non-acetylated

Dilution	Expected (pmol/mL)	Observed (pmol/mL)	Recovery (%)
Neat	---	45.6	---
1:2	22.8	24.6	108%
1:4	11.4	13.5	118%
1:8	5.7	6.3	111%
1:16	2.9	3.0	103%
1:32	1.4	1.9	135%
1:64	0.71	0.96	135%
1:128	0.36	0.38	106%

Acetylated

Dilution	Expected (pmol/mL)	Observed (pmol/mL)	Recovery (%)
Neat	---	3.4	---
1:2	1.7	1.7	100%
1:4	0.85	0.89	105%
1:8	0.43	0.42	98%
1:16	0.21	0.19	90%

A 0.1M HCl sample containing cGMP was serially diluted 1:2 in the 0.1M HCl diluent and measured in the assay. The results are shown in the table below.

Non-acetylated

Dilution	Expected (pmol/mL)	Observed (pmol/mL)	Recovery (%)
Neat	---	87.4	---
1:2	43.7	48.8	112%
1:4	21.9	25.4	116%
1:8	10.9	10.1	93%
1:16	5.5	6.3	115%
1:32	2.7	3.3	122%

Acetylated

Dilution	Expected (pmol/mL)	Observed (pmol/mL)	Recovery (%)
Neat	---	14.4	---
1:2	7.2	9.4	130%
1:4	3.6	4.4	122%
1:8	1.8	2.2	122%
1:16	0.90	1.3	144%
1:32	0.45	0.63	140%
1:64	0.23	0.22	96%

Precision

Intra-assay precision was determined by assaying 20 replicates of three buffer controls containing cGMP in a single assay.

Non-Acetylated Format

In Assay Buffer 2		In 0.1M HCl	
pmol/mL	%CV	pmol/mL	%CV
1.5	5.2	1.6	4.4
16.6	4.0	9.9	7.9
481	7.6	115	6.6

Acetylated Format

In Assay Buffer 2		In 0.1M HCl	
pmol/mL	%CV	pmol/mL	%CV
0.54	6.5	0.58	9.6
1.5	4.6	1.4	3.6
6.8	4.5	5.4	3.5

Inter-assay precision was determined by measuring buffer controls of varying cGMP concentrations in multiple assays over several days.

Non-Acetylated Format

In Assay Buffer 2		In 0.1M HCl	
pmol/mL	%CV	pmol/mL	%CV
1.8	13.7	2.1	6.0
16.9	3.5	8.5	9.9
359	5.0	92	6.9

Acetylated Format

In Assay Buffer 2		In 0.1M HCl	
pmol/mL	%CV	pmol/mL	%CV
0.70	5.9	0.35	11
2.0	6.2	3.6	8.4
8.6	6.8	10	4.6

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Product Manual

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