



cGMP ELISA kit
Catalog No. ADI-900-013
96 Well Kit

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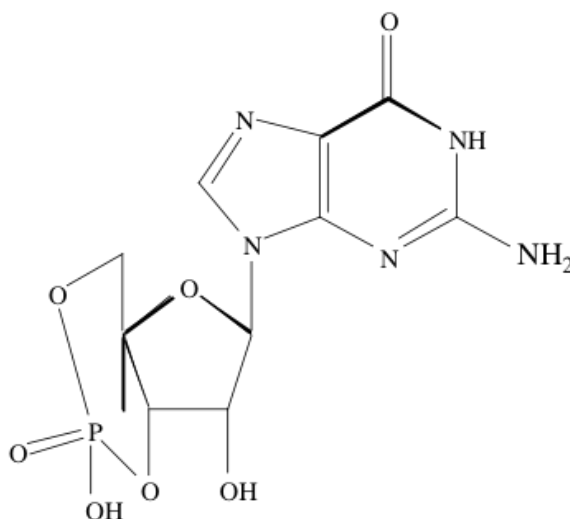
Description

The cyclic GMP Enzyme-Linked Immunosorbent Assay (ELISA) kit is a competitive immunoassay for the quantitative determination of cyclic GMP in samples. The kit uses a polyclonal antibody to cGMP to bind, in a competitive manner, the cGMP in the standard or sample or an alkaline phosphatase molecule which has cGMP covalently attached to it. After a simultaneous incubation at room temperature the excess reagents are washed away and substrate is added. After a short incubation time the enzyme reaction is stopped and the yellow color generated read on a microplate reader at 405 nm. The intensity of the bound yellow color is inversely proportional to the concentration of cGMP in either standards or samples. The measured optical density is used to calculate the concentration of cGMP. For further explanation of the principles and practice of immunoassays please see the excellent books by Chard¹ or Tijssen².

Introduction

Guanosine 3', 5'-cyclic monophosphate (cyclic GMP; cGMP) was identified in 1963³. It has been shown to be present at levels typically 10-100 fold lower than cAMP in most tissues and is formed by the action of the enzyme guanylate cyclase on GTP. It is involved in a number of important biological reactions. Some hormones, such as acetylcholine, insulin and oxytocin, as well as certain other chemicals like serotonin and histamine, cause an increase in cGMP levels^{4,5}. Stimulators of guanylate cyclase such as the vasodilators nitroprusside, nitroglycerin, sodium nitrate and nitric oxide (NO) also stimulate cGMP levels⁶. Peptides, such as atrial natriuretic peptide (ANP) that relax smooth muscle also increase cGMP concentrations⁷. cGMP has been confirmed as a second messenger for ANP⁸. NO can be synthesized from L-arginine and diffuse through cell membranes^{9,10}. The interaction of NO with guanylate cyclase allows cGMP to act as a third messenger in some cells¹¹.

Cyclic GMP



Precautions

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1. Some kit components contain azide, which may react with lead or copper plumbing. When disposing of reagents always flush with large volumes of water to prevent azide build-up.
2. Some solutions supplied in this kit are caustic; care should be taken with their use.
3. The activity of the alkaline phosphatase conjugate is dependent on the presence of Mg^{2+} and Zn^{2+} ions. The activity of the conjugate is affected by concentrations of chelators (>10 mM) such as EDTA and EGTA.
4. We tested this kit's performance with a variety of samples, however it is possible that high levels of interfering substances may cause variation in assay results.
5. The cyclic GMP Standard provided, Catalog No. 80-0153, is supplied in ethanolic buffer at a pH optimized to maintain cGMP integrity. Care should be taken in handling this material because of the known and unknown effects of cGMP.

CAUTION: Some components of this Kit contain chemicals that are lachrymators, corrosive and flammable. Use with caution and wear suitable protection. Refer to the Material Safety Data Sheets available on our Web site or by fax.

Materials Supplied

- 1. Goat anti-Rabbit IgG Microtiter Plate, One Plate of 96 Wells, Catalog No. 80-0060**
A plate using break-apart strips coated with goat antibody specific to rabbit IgG.
- 2. cGMP EIA Conjugate, 5 mL, Catalog No. 80-0151**
A blue solution of alkaline phosphatase conjugated with cGMP.
- 3. cGMP EIA Antibody, 5 mL, Catalog No. 80-0152**
A yellow solution of a rabbit polyclonal antibody to cGMP.
- 4. Assay Buffer 2, 27 mL, Catalog No. 80-0055**
A buffer containing proteins and sodium azide as preservative.
- 5. Wash Buffer Concentrate, 27 mL, Catalog No. 80-1286**
Tris buffered saline containing detergents.
- 6. Cyclic GMP Standard, 0.5 mL, Catalog No. 80-0153**
A solution of 5,000 pmol/mL cGMP.
- 7. p-Npp Substrate, 20 mL, Catalog No. 80-0075**
A solution of p-nitrophenyl phosphate in buffer. Ready to use.
- 8. Stop Solution, 5 mL, Catalog No. 80-0247**
A solution of trisodium phosphate in water. Keep tightly capped.
Caution: **Caustic.**
- 9. Acetylation Kit, 2 vials, Catalog No. 950-001**
 - a. Triethylamine, 2 mL, Catalog No. 80-0063.**
CAUTION: Lachrymator, Harmful Vapor, Flammable.
 - b. Acetic Anhydride, 1 mL, Catalog No. 80-0064.**
CAUTION: Lachrymator, Corrosive, Flammable.
- 10. cGMP Assay Layout Sheet, 1 each, Catalog No. 30-0030**
- 11. Plate Sealer, 1 each, Catalog No. 30-0012**

Storage

All components of this kit, except the standard and conjugate, are stable at 4°C until the kit's expiration date. Upon receipt, store the standard and conjugate at -20°C. Recommended storage temperatures do not necessarily reflect shipping conditions.

Materials Needed but Not Supplied

1. Deionized or distilled water.
2. Precision pipets for volumes between 5 µL and 1,000 µL.
3. Repeater pipets for dispensing 50 µL and 200 µL.
4. Disposable beakers for diluting buffer concentrates.
5. Graduated cylinders.
6. A microplate shaker.
7. Adsorbent paper for blotting.
8. Microplate reader capable of reading at 405 nm, preferably with correction between 570 and 590 nm.

Sample Handling

The cGMP ELISA kit is compatible with cGMP samples in a wide range of matrices. Samples diluted sufficiently into Assay Buffer 2 can be read directly from the standard curve. Please refer to the Sample Recovery recommendations on page 14 for details of suggested dilutions. However, the end user **must verify** that the recommended dilutions are appropriate for their samples. **Samples containing rabbit IgG may interfere with the assay.** If samples with very low levels of cGMP are to be measured we have provided reagents to acetylate samples and standards. Please refer to References 12-18 for further methods of extraction of cGMP from samples.

Samples in the majority of tissue culture media, including those containing fetal bovine serum, can also be read in the assay, provided the standards have been diluted into the tissue culture media instead of Assay Buffer 2. There will be a small change in the binding associated with running the standards and samples in media. Users should only use standard curves generated in media or buffer to calculate concentrations of cGMP in the appropriate matrix.

Urine samples may be used in the assay directly after dilution in Assay Buffer 2. Plasma samples should be drawn in tubes containing EDTA. EDTA chelates calcium and will stop phosphodiesterase activity. The plasma collected should be assayed immediately or frozen below -20°C. Plasma samples can be diluted with Assay Buffer 2 per recommendation on plate 12. If samples cannot be diluted, the following extraction method may be used. (Add 2 mL of 95% ethanol to 1 mL of the collected plasma. Vortex for 15 seconds and let sit at room temperature for 5 minutes. Centrifuge

for 10 mins. at 600 x g at room temperature. Decant the supernatant into a clean tube. These samples should be dried down, reconstituted in Assay Buffer 2 and then used directly in the assay.)

Tissue samples should be rapidly frozen in liquid nitrogen. Grind the frozen tissue to a fine powder under liquid nitrogen in a stainless steel mortar. Weigh the frozen tissue and homogenize in 10 volumes of cold 5% TCA in a glass-Teflon tissue grinder. Centrifuge at 600 x g for 10 minutes. Extract the supernatants with 3 volumes of water-saturated ether. Dry the aqueous extracts and run the reconstituted samples directly in the assay.

For cells adhering to glass or plastic, use our Direct cGMP kit, Catalog Numbers ADI-900-014 or ADI-901-014.

Procedural Notes

1. Do not mix components from different kit lots or use reagents beyond the kit expiration date.
2. Allow all reagents to warm to room temperature for at least 30 minutes before opening.
3. Standards can be made up in either glass or polypropylene tubes; avoid polystyrene.
4. Pre-rinse the pipet tip and use fresh tips for each sample, standard and reagent.
5. Pipet standards and samples to the bottom of the wells.
6. Add the reagents to the side of the well to avoid contamination.
7. This kit uses break-apart microtiter strips, which allow the user to measure as many samples as desired. Unused wells must be kept desiccated at 4°C in the sealed bag provided. The wells should be used in the frame provided.
8. Care must be taken to minimize contamination by endogenous alkaline phosphatase. Contaminating alkaline phosphatase activity, especially in the substrate solution, may lead to high blanks. Care should be taken not to touch pipet tips and other items that are used in the assay with bare hands.
9. **Prior to addition of substrate, ensure that there is no residual wash buffer in the wells. Any remaining wash buffer may cause variation in assay results.**

Reagent Preparation

1. cGMP Standard - Non-Acetylated Version

Allow the 5,000 pmol/mL cGMP standard solution to warm to room temperature. Label six 12 x 75 mm glass tubes #1 through #6. Pipet 900 μ L standard diluent (Assay Buffer 2 or Tissue Culture Media) into tube #1 and 800 μ L of standard diluent into tubes #2-6. Add 100 μ L of the 5,000 pmol/mL standard to tube #1. Vortex thoroughly. Add 200 μ L of tube #1 to tube #2 and vortex thoroughly. Continue this for tubes #3 through #6. **The concentration of cGMP in tubes #1 through #6 will be 500, 100, 20, 4, 0.8 and 0.16 pmol/mL respectively. See cGMP Assay Layout Sheet for dilution details. Diluted standards should be used within 60 minutes of preparation.**

2. Acetylation Reagent

Prepare the Acetylating Reagent by adding 0.5 mL of acetic anhydride to 1 mL triethylamine. **Use the prepared reagent within 60 minutes of preparation.**

3. cGMP Standard - Acetylated Version

Allow the 5,000 pmol/mL cGMP standard solution to warm to room temperature. Label five 12 x 75 mm glass tubes #1 through #5. Pipet 980 μ L standard diluent (Assay Buffer 2 or Tissue Culture Media) into tube #1 and 800 μ L standard diluent into tubes #2-5. Add 20 μ L of the 5,000 pmol/mL standard to tube #1. Vortex thoroughly. Add 200 μ L of tube #1 to tube #2 and vortex thoroughly. Continue this for tubes #3 through #5.

The concentration of cGMP in tubes #1 through #5 will be 100, 20, 4, 0.8, and 0.16 pmol/mL respectively. See cGMP Assay Layout Sheet for dilution details.

Acetylate all standards and samples by adding 10 μ L of the Acetylating Reagent for each 200 μ L of standard or sample. Add the reagent directly to the samples and vortex for 2 seconds.

Label one 12 x 17 mm glass tube as the Zero Standard/NSB tube. Pipet 1 mL standard diluent into this tube. Add 50 μ L of the Acetylating Reagent to the Zero Standard/NSB tube and use in Step 2 and Step 5 in the Assay Procedure.

Failure to acetylate the NSB and Zero standard will result in inaccurate B/Bo values. Use the acetylated standards or samples within 30 minutes.

4. Wash Buffer

Prepare the Wash Buffer by diluting 5 mL of the supplied concentrate with 95 mL of deionized water. This can be stored at room temperature until the kit expiration, or for 3 months, whichever is earlier.

Assay Procedure

Bring all reagents to room temperature for at least 30 minutes prior to opening.

All standards and samples should be run in duplicate.

If Acetylated Version of the kit is to be run, acetylate all standards and samples by adding 10 μL of the Acetylating Reagent for each 200 μL of standard or sample. Add 50 μL of the Acetylating Reagent to the Zero Standard/NSB tube (Refer to Step 3 in Reagent Preparation section) and use in Steps 2 and 5 below (failure to acetylate the NSB and Zero standard will result in inaccurate B/Bo values). Add the reagent directly to the samples and vortex for 2 seconds. Use the acetylated standards or samples within 30 minutes.

1. Refer to the Assay Layout Sheet to determine the number of wells to be used and put any remaining wells with the desiccant back into the pouch and seal the ziploc. Store unused wells at 4°C.
2. Pipet 100 μL of standard diluent (Assay Buffer 2 or Tissue Culture Media) into the NSB and the Bo (0 pmol/mL Standard) wells.
3. Pipet 100 μL of Standards into the appropriate wells.
4. Pipet 100 μL of the Samples into the appropriate wells.
5. Pipet 50 μL of Standard Diluent into the NSB wells.
6. Pipet 50 μL of blue Conjugate into each well except the TA and Blank wells.
7. Pipet 50 μL of yellow Antibody into each well, except the Blank, TA and NSB wells.

NOTE: Every well used should be **Green** in color except the NSB wells which should be **Blue**. The Blank and TA wells are empty at this point and have no color.

8. Incubate the plate at room temperature for 2 hours on a plate shaker at ~500 rpm. The plate may be covered with the plate sealer provided, if so desired. **If using the Acetylated overnight format, incubate for 18-24 hours at 4°C.**
9. Empty the contents of the wells and wash by adding 400 μL of wash solution to every well. Repeat the wash 2 more times for a total of **3 washes**.
10. After the final wash, empty or aspirate the wells, and firmly tap the plate on a lint free paper towel to remove any remaining wash buffer.
11. Add 5 μL of the blue Conjugate to the TA wells.

12. Add 200 μ L of the pNpp Substrate solution to every well. Incubate at room temperature for 1 hour without shaking.
13. Add 50 μ L of Stop Solution to every well. This stops the reaction and the plate should be read immediately.
14. Blank the plate reader against the Blank wells, read the optical density at 405 nm, preferably with correction between 570 and 590 nm. If the plate reader is not able to be blanked against the Blank wells, manually subtract the mean optical density of the blank wells from all readings.

Calculation of Results

Several options are available for the calculation of the concentration of cGMP in the samples. We recommend that the data be handled by an immunoassay software package utilizing a weighted 4 parameter logistic curve fitting program. If this type of data reduction software is not readily available, the concentration of cGMP can be calculated as follows:

1. Calculate the average Net Optical Density (OD) bound for each standard and sample by subtracting the average NSB OD from the average OD bound:

$$\text{Average Net OD} = \text{Average Bound OD} - \text{Average NSB OD}$$

2. Calculate the binding of each pair of standard wells as a percentage of the maximum binding wells (Bo), using the following formula:

$$\text{Percent Bound} = \frac{\text{Net OD}}{\text{Net Bo OD}} \times 100$$

3. Using Logit-Log paper plot Percent Bound (B/Bo) versus Concentration of cGMP for the standards. Approximate a straight line through the points. The concentration of cGMP in the unknowns can be determined by interpolation.

Samples with concentrations outside of the standard curve range will need to be re-analyzed using a different dilution.

To normalize for protein content, divide the resulting picomole per mL determinations (pmol/mL) by the total protein concentration (mg/mL) in each sample. This is expressed as pmol cGMP per mg of total protein.

Typical Results

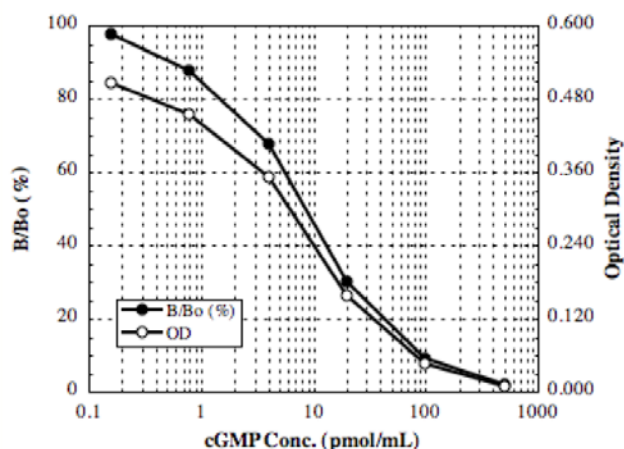
The results shown below are for illustration only and **should not** be used to calculate results from another assay.

<u>Average Sample</u>	<u>Non-Acetylated Version</u>			<u>Acetylated Version</u>		
	<u>Percent Net OD</u>	<u>cGMP Bound</u>	<u>(pmol/mL)</u>	<u>Percent Net OD</u>	<u>cGMP Bound</u>	<u>(pmol/mL)</u>
Blank OD	(0.176)			(0.146)		
TA	0.296			0.320		
NSB	0.000	0%		0.002	0%	
Bo	0.519	100%	0	0.266	100%	0
S1	0.010	1.9%	500	0.003	1.1%	100
S2	0.047	9.1%	100	0.018	6.8%	20
S3	0.156	30.1%	20	0.062	23.3%	4
S4	0.352	67.8%	4	0.149	56.0%	0.8
S5	0.455	87.7%	0.8	0.223	83.8%	0.16
S6	0.507	97.7%	0.16	--	--	--
Unknown 1	0.300	58.9%	6.05	0.130	48.9%	1.12
Unknown 2	0.098	18.9%	39.3	0.010	3.8%	33.9

Typical Standard Curves

Typical standard curves are shown below. These curves **must not** be used to calculate cGMP concentrations; each user must run a standard curve for each assay and version used.

Non-Acetylated Version

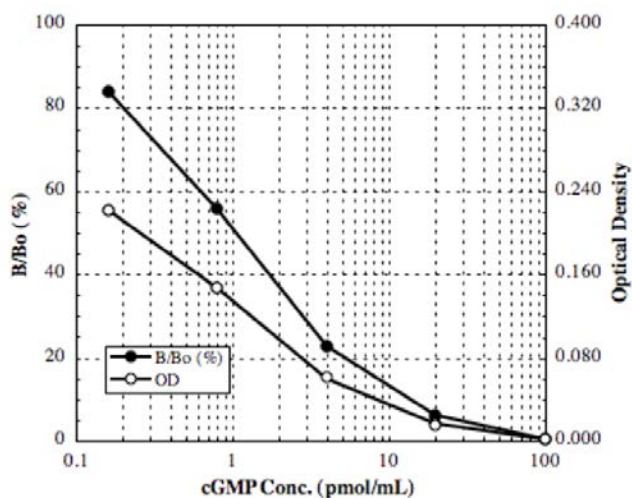


Typical Quality Control Parameters

Total Activity Added = $0.296 \times 10 = 2.96$
 %NSB = 0.0%
 %Bo/TA = 17.6%
 Quality of Fit = 0.9999

20% Intercept = 36.5 pmol/mL
 50% Intercept = 8.4 pmol/mL
 80% Intercept = 1.9 pmol/mL

Acetylated Version



Typical Quality Control Parameters

Total Activity Added = $0.320 \times 10 = 3.20$
 %NSB = 0.1%
 %Bo/TA = 8.3%
 Quality of Fit = 1.0000

20% Intercept = 5.1 pmol/mL
 50% Intercept = 1.1 pmol/mL
 80% Intercept = 0.2 pmol/mL

Performance Characteristics

The following parameters for this kit were determined using the guidelines listed in the National Committee for Clinical Laboratory Standards (NCCLS) Evaluation Protocols¹⁹.

Sensitivity

Sensitivity was calculated by determining the average optical density bound for sixteen (16) wells run with the Bo, and comparing to the average optical density for sixteen (16) wells run with Standard #6 in the Non-Acetylated, or with Standard #5 in the Acetylated version. The detection limit was determined as the concentration of cGMP measured at two (2) standard deviations from the zero along the standard curve.

Non-Acetylated Version

Mean OD for Bo =	0.397 ± 0.013 (3.3%)
Mean OD for Standard #6 =	0.385 ± 0.012 (3.2%)
Delta Optical Density (0-0.16 pmol/mL) = 0.397 - 0.385 =	0.012
2 SD's of the zero standard =	0.026
Sensitivity = $\frac{0.026}{0.012} \times 0.16 \text{ pmol/mL} =$	0.37 pmol/mL

Acetylated Version

Mean OD for Bo =	0.280 ± 0.008 (2.8%)
Mean OD for Standard #5 =	0.251 ± 0.013 (5.3%).
Delta Optical Density (0-0.16 pmol/mL) = 0.280 - 0.251 =	0.029
2SD's of the zero standard =	0.016
Sensitivity = $\frac{0.016}{0.029} \times 0.16 \text{ pmol/mL} =$	0.088 pmol/mL

Linearity

Non-Acetylated Version

A sample containing 800 pmol/mL cGMP was serially diluted 8 times 1:2 in the kit Assay Buffer 2 and measured in the assay. The data was plotted graphically as actual cGMP concentration versus measured cGMP concentration.

The line obtained had a slope of 1.060 with a correlation coefficient of 0.998.

Acetylated Version

A sample containing 5.79 pmol/mL cGMP was serially diluted 3 times 1:2 in the kit Assay Buffer 2 and measured in the Acetylated version of the assay. The data was plotted graphically as actual cGMP concentration versus measured cGMP concentration.

The line obtained had a slope of 0.903 with a correlation coefficient of 0.999.

Precision

Intra-assay precision was determined by taking samples containing low, medium and high concentrations of cGMP and running these samples multiple times (n=16) in the same assay. Inter-assay precision was determined by measuring three samples with low, medium and high concentrations of cGMP in multiple assays (n=16).

The precision numbers listed below represent the percent coefficient of variation for the concentrations of cGMP determined in these assays as calculated by a 4 parameter logistic curve fitting program.

	<u>Non-Acetylated Version</u>			<u>Acetylated Version</u>		
	cGMP	Intra-assay	Inter-assay	cGMP	Intra-assay	Inter-assay
	<u>(pmol/mL)</u>	<u>(%CV)</u>	<u>(%CV)</u>	<u>(pmol/mL)</u>	<u>(%CV)</u>	<u>(%CV)</u>
Low	1.5	5.2		0.54	6.5	
Medium	16.6	4.0		1.50	4.6	
High	481	7.6		6.81	4.5	
Low	1.8		13.7	0.70		5.9
Medium	16.9		3.5	1.96		6.2
High	359		5.0	8.61		6.8

Cross Reactivities

The cross reactivities for a number of related compounds was determined by dissolving the cross reactant (purity checked by N.M.R. and other analytical methods) in Assay Buffer 2 at concentrations from 10,000 to 10 pmol/mL. These samples were then measured in the cGMP assay, and the measured cGMP concentration at 50% B/Bo calculated. The % cross reactivity was calculated by comparison with the actual concentration of cross reactant in the sample and expressed as a percentage.

<u>Compound</u>	<u>Cross Reactivity</u>
cGMP	100%
GMP	<0.001%
GTP	<0.001%
cAMP	<0.001%
AMP	<0.001%
ATP	<0.001%
cUMP	<0.001%
CTP	<0.001%

Sample Recoveries

Please refer to pages 4 and 5 for Sample Handling recommendations and Standard preparation.

cGMP concentrations were measured in a variety of different samples including tissue culture media, human saliva, serum, and urine. cGMP was spiked into the undiluted samples which were diluted with the appropriate diluent and then assayed in the kit. The following results were obtained:

<u>Sample</u>	<u>Non-Acetylated Version</u>		<u>Acetylated Version</u>	
	<u>% Recovery</u>	<u>Recommended Dilution*</u>	<u>% Recovery</u>	<u>Recommended Dilution*</u>
Tissue Culture Media	101.7	1:100	95.8	Undiluted
Human Saliva	102.9	1:10		
Human Serum	101.3	1:10-1:100		Not Recommended
Human Heparin Plasma	104.4	1:10		
Human EDTA Plasma	115.0	1:10-1:100	93.6	≥ 1:2
Human Urine	97.7	1:100-1:1000		

* See Sample Handling instructions on page 5 for details.

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